

## Whey Protein Supplementation Improves Nutritional Status, Glutathione Levels, and Immune Function in Cancer Patients: A Randomized, Double-Blind Controlled Trial

Akkarach Bumrungpert,<sup>1</sup> Patcharanee Pavadhgul,<sup>1</sup> Pornpimon Nunthanawanich,<sup>1</sup>  
Anchalee Sirikanchanarod,<sup>1</sup> and Araya Adulbhan<sup>2</sup>

<sup>1</sup>Department of Nutrition, Faculty of Public Health, Mahidol University, Bangkok, Thailand.

<sup>2</sup>Department of Chemotherapy, National Cancer Institute, Bangkok, Thailand.

**ABSTRACT** Clinical side effects from medical therapy play an important role in causing malnutrition among cancer patients. Whey protein isolates (WPIs) have the potential to improve the nutritional status of cancer patients. The present study determined the effects of whey protein supplementation on nutritional status, glutathione (GSH) levels, immunity, and inflammatory markers in cancer patients in Thailand. A total of 42 cancer patients (41–63 years old) who received intravenous chemotherapy were randomized in a double-blind controlled trial at the National Cancer Institute in Thailand. Patients received 40 g of WPI plus zinc and selenium (intervention group,  $n=23$ ) or a maltodextrin oral snack (control group,  $n=19$ ) every day during the daytime for 12 weeks. Nutritional status, GSH levels, immunity, and inflammatory markers were assessed at baseline, 6, and 12 weeks. Whey protein supplementation significantly increased albumin (2.9%) and immunoglobulin G (4.8%) levels compared to the control group at week 12. Controls showed a significantly lower percent change in GSH levels (6.0%), whereas there was a significant time-dependent increase in the intervention group (11.7%). Whey protein supplementation improved nutrition status scores in the intervention group compared to the control. These data indicate that whey protein supplementation can increase GSH levels and improve nutritional status and immunity in cancer patients undergoing chemotherapy. These results will facilitate implementation of malnutrition risk prevention strategies and improve protein status, including immune function, during chemotherapy.

**KEYWORDS:** • albumin • cancer • glutathione • immunity • whey protein

### INTRODUCTION

CANCER IS A LEADING CAUSE OF DEATH in Thailand. The number of cancer cases has increased from 94.8 to 112.8 per 100,000 people from 2011 to 2015.<sup>1</sup> Importantly, cancer treatments are known to cause serious malnutrition issues.<sup>2</sup> Gastrointestinal symptoms experienced by cancer patients, such as loss of appetite, nausea, vomiting, bloating, constipation, and diarrhea, are related to inadequate energy and protein intake, which is the main cause of malnutrition.<sup>2–4</sup> Thus, nutritional therapy should be applied together with medication to alleviate chemotherapy-related malnutrition, and a high energy and protein diet is recommended for cancer patients by the American Society for Parenteral and Enteral Nutrition.<sup>5</sup>

Whey protein isolate (WPI) is an alternative oral nutrition supplement (ONS) that is suitable for cancer patients due to its lactose- and fat-free composition, high quality of protein with excellent amino acid profiles, and high digestibility.<sup>6</sup>

The benefits of WPI have been shown to include immune system support,<sup>7</sup> improved muscle strength and synthesis, and improved metabolism.<sup>6,7</sup> In terms of immune support, WPI may increase GSH function because cysteine-enriched supplementation can lessen oxidative radical formation and prevent infection.<sup>8</sup>

Trace elements, such as zinc (Zn) and selenium (Se), are key nutrients related to cell membrane protection. Zn is a cofactor of glutathione peroxidase and superoxide dismutase enzymes, both of which have the ability to scavenge free radicals and eliminate oxidants, such as hydrogen peroxide.<sup>7,9</sup> Furthermore, Se can interact with vitamin E and provide antiaging properties by acting as a cellular booster.<sup>10</sup> Thus, WPI enriched with Zn and Se improves cell-mediated immunity and antioxidant capacity, which would be beneficial for cancer patients undergoing chemotherapy.

However, there is a limited amount of research, especially human trials, on the beneficial effect of WPI supplements in cancer patients. Therefore, the purpose of the present study was to determine the effect of supplementation with WPI enriched with Zn and Se on the nutritional status, GSH level, immunity, and inflammatory status in cancer patients who are undergoing chemotherapy in Thailand.

Manuscript received 1 September 2017. Revision accepted 29 January 2018.

Address correspondence to: Akkarach Bumrungpert, PhD, Department of Nutrition, Faculty of Public Health, Mahidol University, 420/1 Rajvithi Road, Ratchathewi, Bangkok 10400, Thailand, E-mail: abnutrition@yahoo.com

## MATERIALS AND METHODS

### Patients and treatments

Forty-eight<sup>11</sup> cancer patients (20–65 years old) receiving intravenous chemotherapy at the National Cancer Institute of Thailand were included in this randomized, double-blind controlled trial. All patients provided written informed consent before study inclusion and agreed to follow the study protocol. All participants were diagnosed by an oncologist as cancer patients without metastatic diseases and able to communicate in Thai. In addition, all patients received their first or second cycle of intravenous chemotherapy without complications and did not plan to have surgery, radiation therapy, or concurrent therapy. Patients were ineligible to take part if they were pregnant or lactating, had underlying kidney or liver diseases, cardiovascular problems, or progression of metastases. Patients who were unable to drink the ONS and/or used herbal medicine or dietary adjuvants, especially those containing whey protein, were also excluded.

Patients were enrolled and randomly assigned using a computer program to the intervention or control group. The intervention group ( $n=24$ ) received 40 g of supplement containing WPI<sup>12</sup> with Zn (2.64 mg/day) and Se (0.76 mg/day), while the control group ( $n=24$ ) was provided with 40 g of placebo (maltodextrin) as a daytime snack. The compositions of supplement are presented in Table 1. The intervention and control groups received the same amount of calories (160 calories) in their snacks, which were provided once a day for 12 weeks, and both types of ONS were similar in texture, color, flavor, and odor. Neither daily overall caloric intake nor protein intake was controlled because we wished to have patients continue their daily lifestyle and dietary patterns. The daily energy intake at baseline for the control and intervention groups was 1144 and 1075 kcal/day, respectively, and after implementation was 1118 and 1265 kcal/day, respectively; baseline protein intake for control and intervention groups was 53 and 48 g/day, respectively, and 63 g (1.06 g/average body weight/day) and 100 g (1.6 g/average body weight/day), respectively, after implementation. Research assistants visited participants to explain the details of this project and measure participant weight, percent body fat, and muscle mass. Ethical approval for the present study was granted by the Ethics Review Committee for Human Research of the Na-

tional Cancer Institute of Thailand (135\_2015RC\_OUT465), and the present study was conducted in accordance with the Declaration of Helsinki on human subjects.

### Biochemical analyses

Blood samples were collected by registered nurses at the beginning of the trial (baseline), week 6, and at the end of the trial (week 12) for the determination of complete blood count, kidney function (blood urea nitrogen and creatinine), liver function (aspartate aminotransferase, alanine aminotransferase, and alkaline phosphatase), protein status (albumin), oxidative stress (GSH), inflammation (high sensitivity C-reactive protein), and immune biomarkers (immunoglobulin G [IgG] and CD4<sup>+</sup>). Albumin levels were measured using the modified bromocresol purple method,<sup>13</sup> and GSH was measured by reduction of 5,5'-dithiobis-(2-nitrobenzoic acid) method.<sup>14</sup> High sensitivity C-reactive protein was determined by latex immunoturbidimetry assay,<sup>15</sup> IgG was measured by enzyme-linked immunosorbent assay,<sup>16</sup> and CD4<sup>+</sup> was determined by flow cytometry.<sup>17</sup> Biochemical assessments were carried out at N Health Asia Lab (Bangkok, Thailand), a medical laboratory with ISO15189:2007 certification. To assess clinical symptoms and quality of life, the Subjective Global Assessment (SGA)<sup>18</sup> was modified to document participant weight patterns within the last 3 months, dietary type, dietary changes, gastrointestinal symptoms, functional capacity, edema, and wasting of subcutaneous fat and muscle. The SGA was graded as follows: grade A, well nourished or low risk of malnutrition (8 points); grade B, suspected or moderate malnutrition (9–16 points); and grade C, severe malnutrition (>16 points). The Thai version of the EORTC QLQ-C30 (version 3)<sup>19</sup> was used to assess participant quality of life. Compliance was checked by asking participants to return supplement packaging and reminding them through telephone every week before their visit to the National Cancer Institute. All patient visits were set on the same day as their physician appointment.

### Statistical analyses

SPSS version 18.0 for Windows was used for statistical analyses. Data that followed a normal distribution are expressed as means  $\pm$  standard deviations. An independent Student's *t*-test was used to evaluate the differences in normally distributed variables between intervention and control groups. The Wilcoxon signed-rank test was used to assess pre- versus post-trial measures within groups. A  $P < .05$  was considered statistically significant.

## RESULTS

General characteristics are presented in Table 2. The dropout rate was  $\sim 12.5\%$  due to cancer progression ( $n=3$ ) and being unable to drink the ONS ( $n=3$ ); thus, a total of 42 patients completed the study. Both groups included males and females, and the average age was 51.5 years in the control group and 54.1 years in the intervention group. There were no significant differences in age, weight, percent weight change, body mass index, muscle mass, percent body fat, complete

TABLE 1. THE COMPOSITIONS OF SUPPLEMENT

Composition	Content
WPI	40 g
$\beta$ -lactoglobulin	27.68 g
$\alpha$ -lactalbumin	5.68 g
IgG	0.84 g
Bovine serum albumin	1.32 g
Glycomacropeptide	0.64 g
Leucine	5.72 g
Isoleucine	2.52 g
Valine	2.24 g
Zn	2.64 mg
Se	0.76 mg

IgG, immunoglobulin G; Se, selenium; WPI, whey protein isolate; Zn, zinc.

TABLE 2. GENERAL CHARACTERISTICS AND BLOOD CHEMISTRY OF PARTICIPANTS

General characteristics and biochemical parameters	Control (n=19)			Whey protein (n=23)			P
	Baseline	6 week	12 week	Baseline	6 week	12 week	
Age (year)	51.5±9.6	—	—	54.1±9.3	—	—	.38
Sex, n (%)							
Male	3 (15.8)	—	—	7 (30.4)	—	—	
Female	16 (84.2)	—	—	16 (69.6)	—	—	
Type of cancer, n (%)							
Breast	13 (68.4)	—	—	13 (56.5)	—	—	
Colon	4 (21.1)	—	—	5 (21.7)	—	—	
Lung	0	—	—	2 (8.7)	—	—	
Rectum	0	—	—	1 (4.3)	—	—	
Stomach	0	—	—	1 (4.3)	—	—	
Cholangiocarcinoma	1 (5.3)	—	—	1 (4.3)	—	—	
Pancreatic	1 (5.3)	—	—	0	—	—	
Lymphoma	0	—	—	0	—	—	
Weight (kg)	58.4±10.8 <sup>a</sup>	59.1±10.7 <sup>a</sup>	59.0±10.5 <sup>a</sup>	62.3±13.5 <sup>a</sup>	63.5±12.6 <sup>a</sup>	63.0±12.3 <sup>a</sup>	.32
Weight change (%)	—	1.3±5.2 <sup>a</sup>	1.2±6.4 <sup>a</sup>	—	1.1±2.2 <sup>a</sup>	1.2±2.9 <sup>a</sup>	
Body mass index (kg/m <sup>2</sup> )	23.6±3.7 <sup>a</sup>	23.6±3.9 <sup>a</sup>	23.6±4.0 <sup>a</sup>	24.9±5.7 <sup>a</sup>	24.5±5.3 <sup>a</sup>	24.6±5.3 <sup>a</sup>	.38
Muscle (kg)	38.3±6.2 <sup>a</sup>	38.9±7.1 <sup>a</sup>	38.7±6.2 <sup>a</sup>	39.6±6.9 <sup>a</sup>	40.0±7.2 <sup>a</sup>	40.1±7.1 <sup>a</sup>	.53
Body fat (%)	30.1±8.9 <sup>a</sup>	30.2±10.5 <sup>a</sup>	29.6±9.6 <sup>a</sup>	31.0±12.2 <sup>a</sup>	29.0±11.9 <sup>a</sup>	29.2±12.4 <sup>a</sup>	.78
Hb (g/dL)	11.8±1.4 <sup>a</sup>	11.8±1.1 <sup>a</sup>	12.0±1.0 <sup>a</sup>	11.3±1.4 <sup>a</sup>	11.0±1.3 <sup>a</sup>	10.9±1.6 <sup>a</sup>	.26
Hct (%)	35.9±3.7 <sup>a</sup>	36.3±3.5 <sup>a</sup>	37.0±3.5 <sup>a</sup>	35.0±4.0 <sup>a</sup>	33.8±3.9 <sup>a</sup>	34.1±4.6 <sup>a</sup>	.46
RBC (×10 <sup>6</sup> /mm <sup>3</sup> )	4.5±0.6 <sup>a</sup>	4.5±0.6 <sup>a</sup>	4.4±0.6 <sup>a</sup>	4.4±0.7 <sup>a</sup>	4.4±0.6 <sup>a</sup>	4.2±0.7 <sup>a</sup>	.95
WBC (×10 <sup>3</sup> /mm <sup>3</sup> )	7.3±1.5 <sup>a</sup>	6.6±2.4 <sup>a</sup>	6.8±3.0 <sup>a</sup>	6.9±2.2 <sup>a</sup>	7.0±2.6 <sup>a</sup>	6.8±2.9 <sup>a</sup>	.57
Plt (×10 <sup>3</sup> /mm <sup>3</sup> )	282.1±70.5 <sup>a</sup>	306.6±96.0 <sup>a</sup>	290.3±82.7 <sup>a</sup>	267.2±74.6 <sup>a</sup>	325.8±98.6 <sup>b</sup>	307.7±94.1 <sup>ab</sup>	.52
BUN (mg/dL)	9.0±2.7 <sup>a</sup>	10.1±3.2 <sup>a</sup>	11.0±3.1 <sup>b</sup>	11.2±3.9 <sup>a</sup>	13.5±5.1 <sup>a</sup>	12.7±5.2 <sup>a</sup>	.40
Cr (mg/dL)	0.7±0.2 <sup>a</sup>	0.7±0.2 <sup>a</sup>	0.7±0.2 <sup>a</sup>	0.8±0.3 <sup>a</sup>	0.7±0.2 <sup>a</sup>	0.7±0.2 <sup>a</sup>	.12
AST (U/L)	32.4±34.9 <sup>a</sup>	29.1±14.8 <sup>a</sup>	40.1±42.5 <sup>a</sup>	42.8±47.0 <sup>a</sup>	28.0±12.1 <sup>a</sup>	30.0±13.8 <sup>a</sup>	.44
ALT (U/L)	26.4±43.4 <sup>a</sup>	26.1±17.8 <sup>a</sup>	31.5±23.0 <sup>a</sup>	27.3±21.6 <sup>a</sup>	22.7±10.5 <sup>a</sup>	25.1±19.2 <sup>a</sup>	.93
ALP (U/L)	92.9±82.3 <sup>a</sup>	86.1±45.3 <sup>a</sup>	99.3±69.7 <sup>a</sup>	106.7±96.7 <sup>a</sup>	102.9±59.9 <sup>a</sup>	117.3±118.6 <sup>a</sup>	.63

Values are means ± SD.

P=Comparison of means between the 2 groups at baseline; significant differences at  $P < .05$ .

Means in a row with superscript letters without a common letter differ within group,  $P < .05$ .

ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; BUN, blood urea nitrogen; RBC, red blood cell; WBC, white blood cell.

blood count, or kidney (blood urea nitrogen or creatinine) or liver (aspartate aminotransferase, alanine aminotransferase, or alkaline phosphatase) function between the two groups at baseline. All patients who completed the study were able to follow the study protocol and consume their assigned ONS as planned. Notably, platelet levels significantly increased in a time-dependent manner in the intervention group compared to the control group.

Protein status, GSH level, and immune function are presented in Table 3. Albumin levels significantly increased at week 12 in the intervention group by 2.9%. GSH (biomarker of oxidative stress) was significantly decreased when consistent with cancer chemotherapy. Nevertheless, GSH levels significantly improved after WPI supplementation at week 6 by 6.0% and at week 12 by 11.7% in the intervention group, compared with the control. Regarding immune function, IgG levels significantly decreased after participants received chemotherapy in the control group. There was a significant increase in the percentage of IgG at weeks 6 (0.5%) and 12 (4.8%) in the intervention group, compared to the control, whereas high sensitivity C-reactive protein and CD4<sup>+</sup> were not significantly different.

The outcome of nutritional status and quality of life measurements are shown in Table 4. The SGA score in the control group was not significantly different from the intervention group, but it was significantly lower at weeks 6 and 12 in the intervention group. At the end of the study (week 12), SGA scores were significantly different between the two groups. While there was no significant change in the control group, the SGA grade significantly improved in the intervention group at week 12. Health status and quality of life scores showed the same pattern; they did not significantly change in the control group, while scores for both of these indicators significantly increased in the intervention group.

## DISCUSSION

WPI supplementation with Zn and Se improved the protein status, GSH level, and immune function of cancer patients undergoing chemotherapy by significantly increasing albumin, GSH, and IgG levels after 12 weeks of intervention. Previous studies support the role that whey protein plays in improving albumin levels and immune function. The WPI used herein was enriched with  $\beta$ -lactoglobulin,  $\alpha$ -lactalbumin,

TABLE 3. BIOMARKERS OF PROTEIN STATUS, OXIDATIVE STRESS, INFLAMMATION, AND IMMUNE FUNCTION

Biomarkers	Control (n=19)			Whey protein (n=23)			P
	Baseline	6 week	12 week	Baseline	6 week	12 week	
Albumin (g/dL)	4.3±0.3 <sup>a</sup>	4.3±0.2 <sup>a</sup>	4.3±0.4 <sup>a</sup>	4.1±0.4 <sup>a</sup>	4.2±0.4 <sup>a</sup>	4.4±0.3 <sup>b</sup>	.07
Albumin change (%)	—	0.6±1.7	-0.6±0.7	—	0.2±1.8	2.9±1.5*	
GSH (μmol/L)	20.7±4 <sup>a</sup>	19.4±3.5 <sup>a, b</sup>	18.5±3.7 <sup>b</sup>	22.8±4.4 <sup>a</sup>	23.9±3.7 <sup>b</sup>	25.0±3.4 <sup>b</sup>	.12
GSH change (%)	—	-4.7±14.2	-9.6±13.3	—	6.0±6.7*	11.7±12.6*	
hs-CRP (mg/L)	4.6±6 <sup>a</sup>	5.1±9.4 <sup>a</sup>	6.8±15.3 <sup>a</sup>	7.0±9.5 <sup>a</sup>	6.5±6.6 <sup>a</sup>	5.4±6.4 <sup>a</sup>	.29
hs-CRP change (%)	—	51.9±122.8	14.5±103.3	—	48.5±114.6	23.8±108.4	
IgG (mg/dL)	1439.6±344.1 <sup>a</sup>	1152.7±329.5 <sup>b</sup>	1277±8241.5 <sup>a,b</sup>	1591.3±333.4 <sup>a</sup>	1581.1±301.5 <sup>a</sup>	1658.0±408.8 <sup>a</sup>	.17
IgG change (%)	—	-13.8±11.3	-9.5±20.4	—	0.5±14.1*	4.8±18.1*	
CD4 <sup>+</sup> (%)	38.0±7.1 <sup>a</sup>	37.1±9.9 <sup>a</sup>	36.0±9.2 <sup>a</sup>	39.6±10.0 <sup>a</sup>	37.3±9.9 <sup>a</sup>	38.9±10.8 <sup>a</sup>	.58
CD4 <sup>+</sup> change (%)	—	-2±20	-5.3±18.7	—	-3.7±186	-1.0±13.5	

Values are means±SD.

P=Comparison of means between the 2 groups at baseline; significant differences at  $P<.05$ .

Means in a row with superscript letters without a common letter differ within group,  $P<.05$ .

\*Significant differences between groups at  $P<.05$ .

GSH, glutathione; hs-CRP, high sensitivity C-reactive protein.

IgG, bovine serum albumin, glycomacropeptide, and branched-chain amino acids leucine, isoleucine, and valine. Leucine has been shown to be a protein-synthesis indicator by stimulating translation and initiation pathways, which could prevent muscle loss during malnutrition.<sup>20</sup> Moreover, the high concentration of cysteine and methionine in WPI, together with Zn and Se enrichment, could have also enhanced immune function through intracellular linkages to form GSH,<sup>21</sup> thereby increasing its levels as seen herein. Regarding protein status, serum albumin levels are a key marker of malnutrition status. In the current study, serum albumin levels significantly increased in a time-dependent manner in the intervention group. According to American Society for Parenteral and Enteral Nutrition guidelines for chemotherapy patients, adequate protein intake (1.6 g/body weight/day) can improve malnutrition problems. Moreover, low levels of serum albumin are associated with worse recovery during chemotherapy, and cancer patients with hypoalbuminemia have a poor prognosis of mortality.<sup>22</sup>

In addition, 10–15% of the WPI used is composed of a form of IgG, which has a potential immune modulatory effect

in humans.<sup>23</sup> Chemotherapeutic drugs can deplete platelet levels by permanently damaging bone marrow cells (site of platelet production)<sup>24</sup>; our study showed a similar decrease in platelet levels in the control group. Vitamins C, D, and K have been shown to increase platelet levels through an immunomodulatory function.<sup>25</sup> A low platelet level can delay the cycle of medical treatment, lead to further disease progression, and result in spontaneous bleeding.<sup>26</sup> While supplementation with WPI significantly increased platelet levels herein, the mechanism by which this occurred is unclear. One possibility could be that WPI improves vitamin B<sub>12</sub> and folate absorption, which in turn increase platelet count.<sup>27–29</sup>

Two major strengths of the present study are its randomized, double-blind controlled design and the number of participants, which were much greater than in previous studies.<sup>7,11,29</sup> Even though the type of cancer in each patient was not stratified herein due to time limitations and ability to include cases, the present research was conducted under the care and consideration of physicians and certified dietitians. In the future, comparisons of WPI supplementation on specific cancer types should be evaluated.

TABLE 4. NUTRITION STATUS AND QUALITY OF LIFE

Assessment	Control (n=19)			Whey protein (n=23)			P
	Baseline	6 week	12 week	Baseline	6 week	12 week	
SGA score	9.2±1.4 <sup>a</sup>	9.1±1.2 <sup>a</sup>	9.0±1.2 <sup>a</sup>	10.0±2.3 <sup>a</sup>	8.5±0.9 <sup>b</sup>	8.4±0.9 <sup>bc,*</sup>	.27
SGA grade, n (%)							
Grade A	7 (36.8)	8 (42.1)	8 (42.1)	7 (30.4)	16 (69.6)	19 (82.6)*	
Grade B	12 (63.2)	11 (57.9)	11 (57.9)	16 (69.6)	7 (30.4)	4 (17.4)*	
Grade C	0	0	0	0	0	0	
Health status score	4.7±0.9 <sup>a</sup>	—	4.9±1.1 <sup>a</sup>	4.5±1.2 <sup>a</sup>	—	5.2±1.0 <sup>b</sup>	.73
Quality of life score	4.9±1.3 <sup>a</sup>	—	5.1±0.9 <sup>a</sup>	4.8±1.7 <sup>a</sup>	—	5.5±1.1 <sup>b</sup>	.94

Values are means±SD.

P=Comparison of means between the 2 groups at baseline; significant differences at  $P<.05$ .

Means in a row with superscript letters without a common letter differ within group,  $P<.05$ .

\*Significant differences between groups at  $P<.05$ .

SGA, Subjective Global Assessment.

Based on the current results, we suspect that there are several mechanisms by which WPI supplementation with additional Zn and Se improves the protein status, GSH levels, and immune function, thereby increasing the overall nutritional status of cancer patients receiving chemotherapy. Thus, WPI supplementation and enrichment with essential minerals, such as Zn and Se, may be an alternative route for improving and supporting nutritional status and quality of life in cancer patients undergoing chemotherapy. However, further studies stratifying particular cancer and chemotherapeutic drug types should be conducted to confirm the comprehensive benefit of this dietary strategy.

### ACKNOWLEDGMENTS

The authors thank Dr. Carol Hutchinson, Department of Nutrition, Faculty of Public Health, Mahidol University, for reading and editing the final article. The authors also thank Mega Lifesciences PTY, Ltd., for providing financial support for this study.

### AUTHOR DISCLOSURE STATEMENT

No competing financial interests exist.

### REFERENCES

1. Bureau of Policy and Strategy, Ministry of Public Health: Vital statistics. In: *Public Health Statistics*, Samcharoenpanich Press, Bangkok, 2015, p. 93.
2. Graeff DA, Vogel J, Jager-Wittenaar H, Chua-Hendriks J, Beijer S: Malnutrition in patients with cancer. *Nederl Tijdschr Geneesk* 2011;156:4911.
3. Ruiz RB, Hernández PS: Diet and cancer: Risk factors and epidemiological evidence. *Maturitas* 2014;77:202–208.
4. Hébuterne X, Lemarié E, Michallet M, Montreuil CB, Schneider SM, Goldwasser F: Prevalence of malnutrition and current use of nutrition support in patients with cancer. *JPEN Parenter Enteral Nutr* 2014;38:196–204.
5. August DA, Huhmann MB: ASPEN clinical guidelines: Nutrition support therapy during adult anticancer treatment and in hematopoietic cell transplantation. *JPEN Parenter Enteral Nutr* 2009;33:472–500.
6. Ha E, Zemel MB: Functional properties of whey, whey components, and essential amino acids: Mechanisms underlying health benefits for active people (review). *J Nutr Biochem* 2003;14:251–258.
7. Bounous G: Whey protein concentrate (WPC) and glutathione modulation in cancer treatment. *Altern Med Rev* 2001;6:342.
8. Ripple MO, Henry WF, Rago RP, Wilding G: Pro-oxidant-antioxidant shift induced by androgen treatment of human prostate carcinoma cells. *J Natl Cancer Inst* 1997;89:40–48.
9. Halliwell B: Antioxidant defence mechanisms: From the beginning to the end (of the beginning). *Free Radic Res* 1999;31:261–272.
10. Finley JW: Increased intakes of selenium-enriched foods may benefit human health. *J Sci Food Agric* 2007;87:1620–1629.
11. See D, Mason S, Roshan R: Increased tumor necrosis factor alpha (TNF- $\alpha$ ) and natural killer cell (NK) function using an integrative approach in late stage cancers. *Immunol Invest* 2002;31:137–153.
12. Keri ND: Therapeutic applications of whey protein. *Altern Med Rev* 2004;9:136–156.
13. Ueno T, Hirayama S, Ito M, *et al.*: Albumin concentration determined by the modified bromocresol purple method is superior to that by the bromocresol green method for assessing nutritional status in malnourished patients with inflammation. *Ann Clin Biochem* 2013;50:576–584.
14. Kalpravidh RW, Siritanaratkul N, Insain P, *et al.*: Improvement in oxidative stress and antioxidant parameters in beta-thalassemia/Hb E patients treated with curcuminoids. *Clin Biochem* 2010;43:424–429.
15. Horiuchi Y, Hirayama S, Soda S, *et al.*: Statin therapy reduces inflammatory markers in hypercholesterolemic patients with high baseline levels. *J Atheroscler Thromb* 2010;17:722–729.
16. Odashima NS, Takayanagui OM, Figueiredo JF: Enzyme linked immunosorbent assay (ELISA) for the detection of IgG, IgM, IgE and IgA against *Cysticercus cellulosae* in cerebrospinal fluid of patients with neurocysticercosis. *Arq Neuropsiquiatr* 2002;60:400–405.
17. Barbesti S, Soldini L, Carcelain G, *et al.*: A simplified flow cytometry method of CD4 and CD8 cell counting based on thermoresistant reagents: Implications for large scale monitoring of HIV-infected patients in resource-limited settings. *Cytometry B Clin Cytom* 2005;68:43–51.
18. Pibul K, Techapongsatorn S, Thiengthiantham R, Manomaipiboon A, Trakulhoon V: Nutritional assessment for surgical patients by Bhumibol nutrition triage (BNT) and subjective global assessment (SGA). *Thai J Surg* 2011;32:45–48.
19. Silpakit C, Sirilertrakul S, Jirajarus M, Sirisinha T, Sirachainan E, Ratanatharathorn V: The European organization for research and treatment of cancer quality of life questionnaire (EORTC QLQ-C30): Validation study of the Thai version. *Qual Life Res* 2006;15:167–172.
20. Anthony JC, Anthony TG, Kimball SR, Jefferson LS: Signaling pathways involved in translational control of protein synthesis in skeletal muscle by leucine. *J Nutr* 2001;131:856S–860S.
21. Gold P: The influence of dietary whey protein on tissue glutathione and the diseases of aging. *Clin Invest Med* 1989;12:343–349.
22. Cabrerizo S, Cuadras D, Gomez BF, Artaza AI, Marín CF, Malafarina V: Serum albumin and health in older people: Review and meta-analysis. *Maturitas* 2015;81:17–27.
23. Kulczycki A, Macdermott RP: Bovine IgG and human immune responses: Con a-induced mitogenesis of human mononuclear cells is suppressed by bovine IgG. *Int Arch Allergy Appl Immunol* 1985;77:255–258.
24. Kuter DJ: Managing thrombocytopenia associated with cancer chemotherapy. *Oncology (Williston Park)*. 2015;29:282–294.
25. Devasagayam TP, Tilak JC, Bolor KK, Sane KS, Ghaskadbi SS, Lele RD: Free radicals and antioxidants in human health: Current status and future prospects. *J Assoc Physicians India* 2004;52:794–804.
26. Tmamyam G, Danielyan S, Lambert MP: Chemotherapy induced thrombocytopenia in pediatric oncology. *Crit Rev Oncol Hematol* 2016;99:299–307.
27. Eslami SM, Karandish SM, Marandi, Zand MA: Effects of whey protein supplementation on hematological parameters in healthy young resistance male athletes. *J Appl Sci* 2010;10:991–995.
28. Bockow B, Kaplan TB: Refractory immune thrombocytopenia successfully treated with high-dose vitamin D supplementation and hydroxychloroquine: Two case reports. *J Med Case Rep* 2013;7:91.
29. Luliano L, Colavita AR, Leo R, Partico D, Violi F: Oxygen free radical and platelet activation. *Free Radic Biol Med* 1997;22:999–1006.